Signosis
Innovative Plate Assay Solutions

Mouse G-CSF ELISA

Catalog Number EA-2501 **(For Research Use Only)**

Introduction

Granulocyte colony-stimulating factor (G-CSF) is a colony-stimulating factor hormone. It is produced by a number of different tissues to stimulate the bone marrow to produce granulocytes and stem cells and to stimulate the bone marrow to release them into the blood. It also stimulates the survival, proliferation, differentiation, and function of neutrophil precursors and mature neutrophils, as well as monocytes. G-CSF influences monocyte functions in an anti-inflammatory way. The stimulation of monocytes with G-CSF results in an attenuation of LPS-induced release of IL-1, TNF-α, IL-12 and IL-18.

Principle of the assay

G-CSF ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes rabbit anti-mouse G-CSF for immobilization on the microtiter wells and biotinated rabbit anti-mouse G-CSF antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the G-CSF molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of G-CSF is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

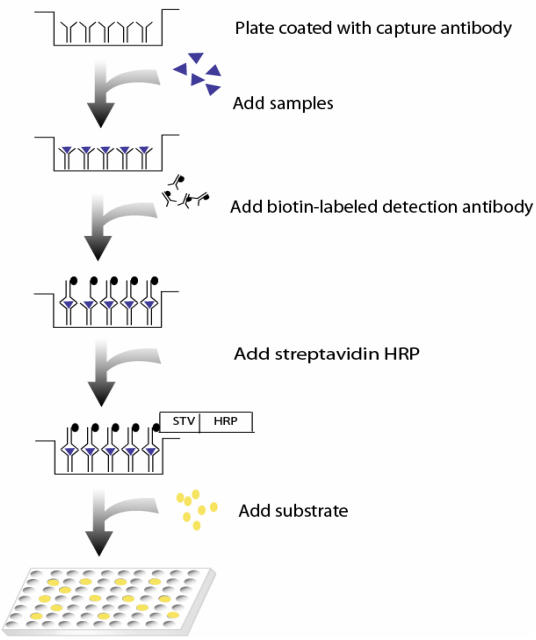


Diagram of ELISA

Materials provided with the kit

- 96 well microplate coated with rabbit anti-mouse G-CSF antibodies (4°C).
- Biotin labeled rabbit anti-mouse G-CSF antibodies (-20°C).
- Streptavidin-HRP conjugate (4°C).
- Recombinant mouse G-CSF standard (200ng/ml) (-20°C).
- 1X Diluent buffer (4°C).
- 5X Assay wash buffer (RT)
- Substrate (4°C).
- Stop Solution (4°C).

Material required but not provided

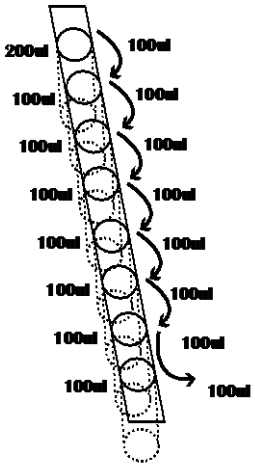
- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer
  40ml 5x Assay wash buffer
  160ml ddH2O
- Dilute 50 times of mouse recombinant G-CSF (200ng/ml) with 1X Diluent buffer to 4000pg/ml and then 2-fold serial dilutions. Add 4ul mouse recombinant G-CSF in 200ul 1X Diluent Buffer (See Step 2 in "Assay Procedure" for detailed instruction)
- Dilute 400 times of biotin labeled rabbit anti-mouse G-CSF antibodies with 1X Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

Assay procedure

1. Cut the sealing film over the plate and remove it from the desired number of well strips. Make sure the rest of wells are well sealed
2. See instruction and diagram below for standard preparation.



a. Add 200ul 1X Diluent buffer to the 1st well. Add 100ul 1X Diluent Buffer to the rest wells of strip.
b. Add appropriate amount of protein recombinant (follow instruction in "Reagent Preparation")
c. Mix dilutions in 1st well and transfer 100ul from the 1st well to the next dilution. (See picture) Incubate each well for 1 hr at room temperature with gentle shaking

3. Add 100ul of sample per well and incubate for 1 hour at room temperature with gentle shaking.
4. Aspirate each well and wash by adding 200µl of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
5. Add 100µl of diluted biotin-labeled mouse anti-human PAI-1 antibody to each well and incubate for 1 hour at room temperature with gentle shaking.
6. Repeat the aspiration/wash as in step 4.
7. Add 100 µl of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.
8. Repeat the aspiration/wash as in step 3.
9. Add 100µl substrate to each well and incubate for 5-30 minutes.
10. Add 50µl of Stop solution to each well. The color in the wells should change from blue to yellow.
11. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes

Example of standard curve